RESEARCH ARTICLE

Open Access

Using informative features in machine learning based method for COVID-19 drug repurposing



Rosa Aghdam^{1*†}, Mahnaz Habibi^{2†} and Golnaz Taheri^{3,4†}

Abstract

Coronavirus disease 2019 (COVID-19) is caused by a novel virus named Severe Acute Respiratory Syndrome Coronavirus-2 (SARS-CoV-2). This virus induced a large number of deaths and millions of confirmed cases worldwide, creating a serious danger to public health. However, there are no specific therapies or drugs available for COVID-19 treatment. While new drug discovery is a long process, repurposing available drugs for COVID-19 can help recognize treatments with known clinical profiles. Computational drug repurposing methods can reduce the cost, time, and risk of drug toxicity. In this work, we build a graph as a COVID-19 related biological network. This network is related to virus targets or their associated biological processes. We select essential proteins in the constructed biological network that lead to a major disruption in the network. Our method from these essential proteins chooses 93 proteins related to COVID-19 pathology. Then, we propose multiple informative features based on drug—target and protein—protein interaction information. Through these informative features, we find five appropriate clusters of drugs that contain some candidates as potential COVID-19 treatments. To evaluate our results, we provide statistical and clinical evidence for our candidate drugs. From our proposed candidate drugs, 80% of them were studied in other studies and clinical trials.

Keywords: Coronavirus disease 2019, SARS-CoV-2, Protein—protein interaction, Clustering method

Introduction

The pandemic situation for Coronavirus disease 2019 (COVID-19) causes more than 197 million infections and more than 4.2 million deaths in more than 200 countries worldwide (until the end of July 2021) and this number is increasing rapidly. Due to this rapid spread, researchers have been searching for therapeutic approaches in the past few months. At present, no medicine has been claimed to be effective in the treatment or even prevention of COIVD-19 [1]. On the other hand, producing new drugs with a complete drug profile is a tough task that

requires extensive time and budget. Drug repurposing is the procedure of perusing new therapeutic uses for available drugs. This process can reduce a large amount of time, money, and danger of the traditional drug discovery process [2]. The main purpose of drug repurposing is to exceed the therapeutic use of the available drugs for more medical scope. Previous researches showed that drugs with similar profiles probably demonstrate similar behavior in the existence of similar targets like proteins [1-3]. Traditional drug repurposing methods are mainly based on finding the relationship between biological activity and the molecular structure of different drugs. However, newer data gathering and analysis shows the urgent need for using computational methods for drug design and repurposing. Computational methods are mainly used to discover different drug interactions that are not considered and found during the clinical trial process [4]. In

Full list of author information is available at the end of the article



© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/loublicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data

^{*}Correspondence: rosa.aghdam@ipm.ir

[†]Rosa Aghdam, Mahnaz Habibi and Golnaz Taheri have contributed equally to this work

¹ School of Biological Sciences, Institute for Research in Fundamental Sciences (IPM), Tehran, Iran

Aghdam et al. J Cheminform (2021) 13:70 Page 2 of 14

drug repositioning, computational methods investigate the relationship between drug databases and genomic, transcriptomic, and other available information with the help of data and network analysis and machine learning methods [2]. Machine-learning based methods for drug repurposing reveal the connection between drugs, viral, and host proteins. In the life cycle of a virus, the viral proteins are associated with different human proteins in the infected cells through different interactions. Within these interactions, the virus hijacks the host cells for replication, and this process changes the regular function of these interacted proteins. Therefore, to design antiviral drugs, a complete understanding of the interaction between human proteins and viral is crucial [5]. It is worth mentioning, in drug repurposing to fight the virus, targeting just virus proteins is not the proper approach. Targeting single virus proteins can cause the viruses to escape this attack through some backup pathways. These backup pathways lead to increased virus resistance with the mutation. Host-directed treatments propose significant strategies [6]. These methods select human proteins as the main carriers for the virus to enter and control human cells. These host-directed treatments seem to be less susceptible to making resistance because human proteins are less influenced by mutations. Therefore, targeting human proteins as drug targets is a more sustainable strategy. In host-directed treatments, it is important to find proteins that are essential for the maintenance and persistence of the disease that is caused by a virus in the human cells. When these proteins are targeted as drug targets, the replication mechanism of the virus collapses. For all of the above-mentioned reasons, repurposing drugs with host-directed treatments against COVID-19 has major potential. Furthermore, drug repurposing methods provide hope for fast practical implementation with the minimum side effects. Molecular interaction and biological interaction networks as valuable resources are the foundation for drug repurposing methods [7]. This means that network-based drug repurposing methods propose novel opportunities for finding drug targets in host-directed treatments [8]. Recent studies show that valuable results are based on viral-host networks for treating HIV [9], Hepatitis C [10], and Ebola as well [11]. Since the outbreak of COVID-19 some research groups have been trying to develop network-based methods to find some repurposed drugs to operate against SARS-CoV-2. Zhou et al. [12] proposed a network-based method for the identification of some candidates as repurposable drugs and some potential drug combinations targeting. Li et al. [13] combined network data with a relative analysis of the gene sequences of the different viruses to find potential drugs for SARS-CoV-2. Gordon et al. [14] proposed a map from human proteins with

SARS-CoV-2 proteins that were found to interact in the affinity purification mass spectrum method. Dick et al. [15] recognized high confidence interactions between human proteins and SARS-CoV-2 proteins with the help of sequence-based protein—protein interaction (PPI) predictors.

In this paper, we propose the four steps method. This method tries to identify novel drug targets and pathways associated with essential proteins in COVID-19. In the first step, we build a graph as a COVID-19 related biological network related to virus targets or their associated biological processes. In the second step, we use two effective algorithms [16, 17] to find the candidate set of proteins from biological networks that lead to a major disruption in the network. In the third step, we identify proteins in our candidate set that are associated with some underlying diseases related to COVID-19. Then, we select 93 proteins as a final set of essential proteins related to disease pathology. Identifying essential proteins may elucidate new drug targets and pathways related to COVID-19. In the fourth and last step, we propose informative features based on drug-protein and PPI networks and find five significant clusters that contain appropriate candidate drugs. Our results show that using our four steps method suggests some appropriate candidate drugs. Most of these candidate drugs are recommended in other studies.

Methods

Finding essential proteins related to COVID-19 pathology as candidate drug targets

Introducing the essential proteins related to COVID-19 pathology as candidate drug targets is one of the most used and appropriate ways to find suitable drugs for COVID-19 treatment. In this subsection, we describe the first, second and third steps of our proposed method. These two steps try to find the set of essential proteins related to COVID-19 pathology. In the first step, we use two effective algorithms [16, 17] for finding the minimum number of proteins that participate in a large number of biological processes. We use these algorithms to find sets of essential proteins based on the disruption of the COVID-19 related biological network. In the second step, we investigated COVID-19 associated protein sets. As a result of this step, we found a subset of essential proteins that are essential to disease pathology.

Construction of COVID-19 related biological network

Suppose that informative biological processes (IBP) is a set of biological processes related to virus targets in COVID-19 that will be described in the next subsection. Two proteins are functionally interacted if they are connected through the same biological processes. A COVID-19

Aghdam et al. J Cheminform (2021) 13:70 Page 3 of 14

related biological network is considered as a weighted undirected graph $G = (V, E, \omega)$. In this graph, each node $v_i \in V$ represents the protein and each edge $e_{ii} \in E$ represents a functional interaction between two nodes v_i and v_i . The $\omega(e_{ij})$ shows the weight of e_{ij} that demonstrates the number of biological processes that two nodes v_i and v_i participate in them. A path between two nodes v_i and v_k in the graph is a sequence of edges that connect the number of distinct nodes through this path. In the weighted graph, the weight of the path between two nodes is defined as follows. Suppose that v_i and v_k as the two ends of this path. Then, the sum of the weight of edges between these two nodes is the weight of this path. A path with the minimum weight between these two nodes is named the shortest path. Now, we define the betweenness value for each node, v_i , in the graph in the following way:

$$Betw(v_i) = \sum_{v_i, v_k \in V} \frac{\theta_{e_{jk}} v_i}{\theta_{e_{jk}}},\tag{1}$$

where $\theta_{e_{jk}}$ shows the total number of shortest paths from node v_j to node v_k and $\theta_{e_{jk}}v_i$ indicates the number of shortest paths that pass through node v_i .

Disruption of COVID-19 related biological processes

We adapt two algorithms to detect the essential proteins in the COVID-19 related biological network [16, 17]. These algorithms [16, 17] select some of the best candidates as removal proteins set from the COVID-19 related biological network to make a major disruption in it. We place the outputs of Algorithm 1 and 2 in Cut_1 and Cut_2 , respectively.

Algorithm 1: spectral partitioning

Partitioning a simple graph, G, into disjoint balanced or nearly balanced parts with removing the minimum number of edges between these two parts is known as the NP-complete problem [16]. We try to approximate this partitioning problem with the spectral partitioning algorithm. This algorithm is based on eigenvectors of the Laplace of the graph, G, and divides the graph into two disjoint parts with respect to eigenvectors of a Laplacian matrix. It is worth mentioning that, the spectral partitioning algorithm is one of the best heuristic approaches for graph partition. Let $A = [a_{ij}]$ shows the adjacency matrix of graph G such that,

$$a_{ij} = \begin{cases} 1 & if(\nu_i, \nu_j) \in E \\ 0 & otherwise \end{cases}$$
 (2)

We define a diagonal degree matrix $D = diag(d_i)$ for graph G. In this matrix value $d(v_i)$ shows the degree of v_i in graph G. The Laplacian matrix of the graph G is defined by $L = D \setminus A$ and $L(G) = [l_{ii}]$ where,

$$l_{ij} = \begin{cases} 1 & if(v_i, v_j) \in E \\ d(v_i) & if i = j \\ 0 & otherwise \end{cases}$$
 (3)

The Laplacian matrix is a symmetric positive semi-definite matrix. This matrix has some important properties. Suppose that vector $u = (u_1, u_2, ..., u_n)$ shows the normalized eigenvectors of matrix L(G) and vector $(\lambda_1, \lambda_2, ..., \lambda_n)$ demonstrates the corresponding eigenvalues of these eigenvectors. We first compute the eigenvectors of Laplacian matrix L(G), according to the second smallest eigenvalue of this matrix $,\lambda_2,$ and put them in vector $X = (x_1, ..., x_n)$. Then, we sort the elements of vector X and insert half of the nodes in partition G_1 and the reminder of nodes in another partition G_2 . This procedure divides the nodes of graph G into two partitions, G_1 and G_2 with nearly equal sizes. Removing the edges between these two parts through the cut edges $E(G_1, G_2)$ makes these two parts disconnect. Suppose the vector $A = \{\alpha_1, ..., \alpha_m\}$ shows the vertices placed in part G_1 and vector $B = \{\beta_1, ..., \beta_m\}$ shows the vertices are placed in part G_2 , respectively. To make these two parts, G_1 and G_2 disconnect, we choose vertices from vectors A and Brepeatedly. The vertices are chosen with respect to their degrees and removed until the all edges in $E(G_1, G_2)$ are covered.

Algorithm 2: betweenness value

This algorithm [17] tries to make the maximum disruption in the network by removing the minimum number of essential proteins. The selection method in algorithm [17] is based on the betweenness value mentioned in Eq 1. The algorithm [17] has three parts. In the first part, the betweenness value for each node in the graph G is calculated. In the second part, to separate the graph G into two disjoint partitions G_1 and G_2 , the node with the minimum betweenness value in graph G is selected and placed in partition G_1 . Then, from all of the neighbors of the selected node, the node with the minimum betweenness value is selected and placed in the other partition G_2 . These procedures are repeated until all nodes are placed into two disjoint partitions G_1 and G_2 . In the third part, the minimum number of nodes from two constructed partitions G_1 and G_2 is selected with respect to their betweenness values to remove all edges in $E(G_1, G_2)$. The third step of this algorithm is equivalent to the minimum bi-section problem that is an *NP*-complete problem [18].

Candidate essential proteins associated with COVID-19 pathology

COVID-19 is a pandemic disease and has different severity and symptoms for various patients. The severity of this disease can vary from asymptomatic to fatal for

Aghdam et al. J Cheminform (2021) 13:70 Page 4 of 14

different people. Recent studies show that this disease has high severity in people with some underlying conditions. Some of the most related underlying diseases are Diabetes, Cardiovascular diseases, Lung diseases, Hepatitis, Kidney disease, and different types of cancer. Hence, we expect that the genetics of these underlying diseases has some correlations with the essential proteins in COVID-19. For finding these essential proteins, we use the relation between gene and disease from Database for Annotation, Visualization, and Integrated Discovery (DAVID). Then, we select some proteins through our two mentioned algorithms that are annotated to four out of five of these specific comorbid diseases. From these selected proteins, proteins with significant p-values as a set of essential proteins associated with COVID-19 are chosen and placed in E as a set of main target candidates of COVID-19 drugs.

Drug clustering method Protein—protein interaction network

We use 5 human high-throughput PPI networks in this work. The first one, Huri, contains 52,248 binary interactions [19]. The second one is collected from the biological general repository for interaction datasets (BioGRID) and contains 296,046 interactions [20]. The BioGRID dataset contains various interactions that are created from different techniques. In this work, we just use the physical interactions between proteins. The three other datasets are human integrated protein-protein interaction reference (HIPPIE) [21], agile protein interactomes dataanalyzer (APID) [22], and homologous interactions (Hint) [23] that contain 57,428, 171,448, and 64,399 experimental interactions, respectively. These interactions are derived from high-throughput yeast-two hybrid (Y2H) and mass spectrometry methods. We map all of the proteins from these five datasets to their corresponding universal protein resource (UniProt) ID [24]. We removed a protein if it could not be mapped to a Uniprot ID. Finally, in this study, we used 25,260 proteins and 304,730 interactions. For all of these proteins, we use biological process terms from gene ontology (GO) term [25] to point out the biological modules in humans. We find that 20,642 proteins from these 25,260 proteins or 81% of them are annotated. We consider a biological process annotation informative if it has these two properties. First, at least *k* proteins are annotated with it. Second, each of its descendant's GO terms needs to have less than k proteins annotated with them. We set 3 as a value of k and we note that 16,021 biological processes corresponding to these 25,260 proteins that are participating in our interactions. We also use 332 human proteins involved in 26 proteins of the SARS-CoV-2 virus that were revealed by Gorden et al. [14]. The set, T, shows these 332 proteins as possible targets of the SARS-CoV-2 virus. For this set of 332 human proteins, we also consider 1374 IBP GO terms as high-confidence SARS-CoV-2 Human PPI.

We define the overlap between two biological processes, p_1 and p_2 in the following way (| . | shows the size):

$$Overlap(p_1, p_2) = \frac{|p_1 \cap p_2|^2}{|p_1||p_2|}.$$
 (4)

Then, the processes with more than 15% overlaps have been removed. Through this filtering method, we have 1213 non-overlapping biological processes corresponding to COVID-19.

Interactive information between drugs and human protein targets

To evaluate our candidate targets, we use all drugs and their corresponding targets interactions that are reported in the UniProt [24]. These interactions contain 6163 drugs from All-Drug group that are reported in UniProt, these drugs have 2898 protein targets. We also use 44 experimental unapproved drugs for COVID-19 that are reported in DrugBank [26]. From these 44 drugs, 27 drugs have no target information and only 17 drugs have the drug target information. These 17 drugs can target 78 proteins in a cell. This group of drugs is denoted as Covid-Drug. The second group of drugs contains 590 drugs as clinical trials for COVID-19. From these 590 drugs, 328 drugs have targets in the PPI network denoted as Clinical-Drug. These 328 drugs can target 888 proteins in a cell.

Construction of drug-target network

We define some topological features in a PPI network for cluster identification of drugs. These features cluster the available experimental unapproved drugs for COVID-19 with respect to these topological properties of their associated targets in the PPI network. To do this, we define a drug—targets network in the following way.

Each drug-targets network is considered as a bipartite graph $H = \langle D, \tau, E^* \rangle$. In graph H, nodes are divided into two different sets. The first one, D, demonstrates the set of experimental unapproved drugs for COVID-19, and the second one, τ , shows the experimental unapproved drug targets. Each edge $e_{vd} \in E^*$ shows that two nodes $v \in \tau$ and $d \in D$ are connected if the node v in a human cell be a target of drug d. In fact τ , contains the proteins that are placed in the intersection of all drug targets with 2898 proteins and set E. Supposed that $G = \langle V, E \rangle$ is a PPI network that contains the set of virus targets (T) and the set of main targets (τ) . Two nodes v_i and v_j are neighbors if there is an edge between them. Suppose that $N(v_i)$ shows a set of all

Aghdam et al. J Cheminform (2021) 13:70 Page 5 of 14

neighbors for a node v_i , therefore $d(v_i) = |N(v_i)|$ indicates the degree of v_i .

We define 3 different informative topological features for each drug, *d*, and its particular targets as follows. The following features are defined with respect to the situation of its main targets and COVID-19 related biological process.

1 $D_T(d)$: The average ratio of the number of neighbors for each protein $v_i \in \tau_d$ that is also placed in set T according to the degree of v_i .

$$D_T(d) = \frac{\sum_{i=1}^{m} \frac{|N(v_i) \cap T|}{d(v_i)}}{|\tau_d|},$$
 (5)

where $\tau_d = \{v_1, v_m\}$ denotes the number of main targets for drug d.

2 The participation rate of τ_d in set π defines as follow:

$$P_{IBP}(d) = 1 - \sum_{p_i \in \pi} \left(\frac{|p_i \cap \tau_d|}{\sum_{p_i \in \pi} |p_i \cap \tau_d|} \right)^2, \quad (6)$$

where set $\pi = \{p_1, p_2, ..., p_k\}$ shows the non-overlapping biological processes corresponding to COVID-19. The possible values for $P_{IBP}(d)$ is between 0 and 1. If the value of $P_{IBP}(d)$ is closer to 1, it means the neighbors of node d have higher distribution in the set of biological processes.

3 $D_P(d)$: The average ratio of the number of neighbors for each protein $v_i \in \tau_d$ that is also placed in set π according to the degree of v_i .

$$D_{P}(d) = \frac{\sum_{i=1}^{m} \frac{|N(v_{i}) \cap P|}{d(v_{i})}}{|\tau_{d}|}.$$
 (7)

where $P = \bigcup_{p_i \in \pi} p_i$.

Clustering method based on topological features of drug targets

Suppose that $G=< V, E, \omega>$ is a COVID-19 related biological network and $H=<\tau, D, E^*>$ is a bipartite drug—target graph $(\tau\subset V)$. Let $E\subset V$ be a set of essential proteins and τ be a set of main targets. Now, for each drug that has at least one target in set τ , we measure the topological features D_P, D_T and P_{IBP} with respect to their targets in τ . Suppose that d^* is a drug from the Covid-Drug group with the corresponding values of topological features $D_P(d^*), D_T(d^*)$ and $P_{IBP}(d^*)$, respectively. According to the near zero threshold ϵ , a drug d is placed in the same cluster with d^* if the following equation is satisfied:

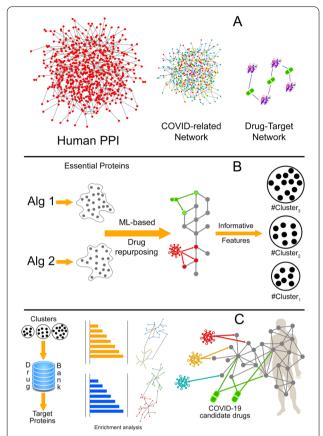


Fig. 1 Overall workflow. Our method integrates a drug–target network with a Human-virus network in the human PPI network. **A**. Human and coronavirus host proteins were collected from different datasets to generate a COVID-19 related biological network. **B**. Algorithm 1 (Alg 1) and 2 (Alg 2) detect the essential proteins in the COVID-19 related biological network. Three Informative features introduced. The Machine Learning method used these features and find three significant clusters. **C**. The resulted clusters evaluated with different measures. These measures are based on drug targets in these clusters. Finally, some candidate drugs recommended

The overall view of our proposed method is illustrated in Fig. 1. Human and coronavirus host proteins were collected from different datasets to generate a COVID-19 related biological network (Part (A)). In Part B, Algorithm 1 (Alg 1) and 2 (Alg 2) are applied to detect the essential proteins in the COVID-19 related biological network. According to the defined features, the clustering method was used to find five appropriate clusters. In Part (C), the resulted clusters evaluated with different measures and some candidate drugs recommended.

$$C = \{d \in D \& |\sqrt{(D_P(d) - D_P(d^*))^2 + (D_T(d) - D_T(d^*))^2 + (P_{IBP}(d) - P_{IBP}(d^*))^2}| < \epsilon\}.$$
(8)

Aghdam et al. J Cheminform (2021) 13:70 Page 6 of 14

Results

Evaluation essential proteins related to COVID-19

The protein sets that are resulted from algorithms 1 and 2 are placed in the sets Cut_1 and Cut_2 , respectively. The union of Cut_1 and Cut_2 is placed in the Cut_{union} set and the intersection of them is placed in $Cut_{intersect}$, respectively. For more evaluation of essential proteins of Cut_1 and Cut_2 sets, we study the topological properties of these two sets. In this work, we claim that through our cut sets as results of two presented algorithms, the maximum number of IBP GO terms are disrupted. We also claim that the two disjoint sets of vertices G_1 and G_2 (resulting from the cut set) are approximately equal in size. Moreover, each IBP GO term, like C has almost the same size on both sides of G_1 and G_2 sets. Suppose that C is a process from the IBP GO terms. The disruption score for this process is defined as follows [16]:

$$Score_{disrupt(C)} = \frac{MAX\{C \cap G_1, C \cap G_2\}}{|C|},$$
 (9)

The closer value of $Score_{disrupt(C)}$ to $\frac{1}{2}$ indicates that process C is completely disrupted. However, if the $Score_{disrupt(C)}$ for a process C is in the range $[0, \frac{1}{2} + \epsilon]$, we say that this process is ϵ -disrupted.

For more evaluation of our proposed cut sets, Cut_1 and Cut_2 , we define two other cut sets with some important topological features. We want to compare our proposed cut sets with these two cut sets and show the advantage of our proposed cut sets with respect to the defined measure $Score_{disrupt(C)}$. In the first cut set,

Table 1 The number of ϵ -disrupted processes for the selected cut sets

	Cut ₁	Cut ₂	Cut _{hub}	Cut _{weight}
0.05-disrupted	745	750	230	298
0.1-disrupted	877	892	378	403
0.2-disrupted	1021	1037	682	733

we select the high degree vertices such that the removal of these vertices divides the graph G into two disjoint parts. This set contains hubs and we named this set as Cut_{hub} . In the second cut set, we select the high weight vertices such that the removal of these vertices divides the graph G into two disjoint parts. We named this set as Cut_{weight} .

In Table 1 we compare the number of ϵ -disrupted processes for Cut_1 , Cut_2 , Cut_{hub} and Cut_{weight} respectively. The results of Table 1 show that Cut_1 and Cut_2 have better disruption properties and this confirms that the selection algorithm that we used for Cut_1 and Cut_2 are significantly better than other algorithms.

In Table 2, we study the number of IBP GO terms that are disrupted through these cut sets. We find that almost all sets disrupt a high number of biological processes. However, Cutunion shows significant superiority with respect to the number of drugs in Covid-Drug and Clinical-Drug groups. Table 2 shows that from these 17 drugs in the Covid-Drug group, 16 drugs are approved with Cutunion, and from 328 drugs in the second group as Clinical-Drug, 273 drugs are approved with Cutunion. Therefore, the set Cutunion could be a possible good candidate to find essential proteins related to COVID-19 pathology as drug targets. To find these essential proteins, we use gene-disease relationship from DAVID tools. We also study some essential proteins in Cutunion that are shared by cardiovascularrelated, hypertension, diabetes type 2, kidney-related and lung-related diseases and placed them in a set E. Table 3 shows 93 proteins of set E that are annotated to four out of five of these specific diseases with significant p-values. We also find that from 17 drugs in the Covid-Drug group, 10 drugs including Bevacizumab, Azithromycin, Ritonavir, Ibuprofen, Colchicine, Darunavir, Methylprednisolone, Tocilizumab, Chloroquine, and Dexamethasone. The results also show that from 328 drugs in Clinical-Drug, 185 drugs are approved by set E. Generally, among 6,163 drugs from the All-Drug

Table 2 The first row shows the number of proteins in sets T, Cut_1 , $Cut_{intersect}$ and Cut_{union} , respectively.

Τ	Cut ₂	Cut ₁	Cut _{intersect}	Cut _{union}
332	2017	2100	1115	3002
1374	1279	1197	1120	1306
1	22	20	15	27
15	218	217	154	281
2	15	15	14	16
30	246	260	225	273
	1374 1 15 2	332 2017 1374 1279 1 22 15 218 2 15	332 2017 2100 1374 1279 1197 1 22 20 15 218 217 2 15 15	332 2017 2100 1115 1374 1279 1197 1120 1 22 20 15 15 218 217 154 2 15 15 14

The number of IBP GO terms overlapped with these subsets collected in the second row. The number of drug targets in each drug group that are associated with these subsets are reported in the third, fourth and fifth rows, respectively. The number of drugs in each drug group that are associated with these subsets are reported in the sixth, seventh and eighth rows, respectively

Aghdam et al. J Cheminform (2021) 13:70 Page 7 of 14

Table 3 Essential protein related to COVID-19 pathology

Essential protein related to COVID-19 pathology

O00206, O14543, O14763, O60603, P00533, P00734, P01019, P01033, P01130, P01133, P01137, P01344 P01374, P01375, P01579, P01583, P01584, P01889, P01891, P01892, P01911, P01912, P02647, P02649 P02751, P02778, P03372, P03989, P04114, P04229, P04439, P04637, P05019, P05089, P05106, P05112 P05164, P05231, P05362, P05534, P06858, P08253, P08571, P08684, P09211, P09601, P10145, P10415 P10635, P11021, P11226, P11473, P13498, P13500, P13501, P14210, P14780, P15692, P16035, P17813 P19438, P19838, P21549, P25445, P28482, P29279, P29459, P29474, P31645, P31749, P35222, P35354 P38936, P40763, P40933, P41597, P42336, P42345, P42898, P48023, P48061, P60568, P78423, P78527 P81172, Q04721, Q14116, Q15848, Q16236, Q30201, Q99958, Q9NR96, Q9Y2R2

group, 1689 drugs are approved by set E. These drugs target 65 proteins out of 93 proteins in set E. We also evaluate set E with respect to the related pathways with DAVID tools. The results show significant pathways related to COVID-19 that have been studied by previous studies [27–30]. A part of these pathways with significant p-values are reported in Table 4.

Annotation cluster 1

Statistical properties of clusters

Enrichment score: 11

As we mentioned earlier, our clustering method introduces 5 different clusters named as #Cluster₁, #Cluster₂, #Cluster₃, #Cluster₄, and #Cluster₅ for 1689 drugs that are approved by proteins in set *E*. Our clustering method uses the defined topological features and 10 approved drugs in Covid-Drug that mentioned in the previous

Table 4 Some of the significantly enriched pathways that are related to COVID-19 essential proteins (E)

Term	Count	P value	
hsa05142:Chagas disease (American trypanosomiasis)	20	2.47E-18	[27]
hsa05323:Rheumatoid arthritis	17	1.79E-15	[27]
hsa05144:Malaria	14	4.85E-15	[27]
hsa05321:Inflammatory bowel disease (IBD)	15	7.61E—15	[27]
hsa05140:Leishmaniasis	14	8.80E-13	[27]
hsa05152:Tuberculosis	19	9.13E—13	[27]
hsa04620:Toll-like receptor signaling pathway	15	1.15E—11	[28]
hsa05146:Amoebiasis	14	1.75E-10	[27]
hsa05145:Toxoplasmosis	14	2.81E-10	[27]
hsa05134:Legionellosis	10	8.56E-09	[29]
hsa05133:Pertussis	10	1.66E-07	[27]
hsa04621:NOD-like receptor signaling pathway	9	2.12E-07	[30]
Annotation cluster 2	Enrichment score:	8.80	
Term	Count	P value	
hsa05168:Herpes simplex infection	20	1.31E—13	[29]
hsa04940:Type I diabetes mellitus	12	8.30E-13	[27]
hsa05332:Graft-versus-host disease	11	1.89E-12	
hsa05330:Allograft rejection	10	2.35E-10	[27]
hsa05320:Autoimmune thyroid disease	7	2.72E-05	[27]
Annotation cluster 3	Enrichment score:	4.53	
Term	Count	P value	
hsa04931:Insulin resistance	10	3.80E-06	
hsa04920:Adipocytokine signaling pathway	8	1.47E-05	[28]
hsa05221:Acute myeloid leukemia	6	4.50E-04	

Aghdam et al. J Cheminform (2021) 13:70 Page 8 of 14

Table 5 The exceeding values (EV) for #Cluster₁ ,#Cluster₂, #Cluster₃ ,#Cluster₄ and #Cluster₅

	EV values
#Cluster ₁	1E-05
#Cluster ₂	0.00018
#Cluster ₃	0.00109
#Cluster ₄	0.0011
#Cluster ₅	0.0013

Table 6 The number of All-Drugs, Covid-Drug and Clinical-Drug for #Cluster₁, #Cluster₂, #Cluster₃, #Cluster₄ and #Cluster₅

	#Cluster ₁	#Cluster ₂	#Cluster ₃	#Cluster4	#Cluster5
No. All-Drugs	2	8	10	28	14
No. Covid-Drug	1	1	1	1	6
No. Clinical- Drug	1	2	3	1	8

subsection, as clustering criteria. In order to evaluate #Cluster₁, #Cluster₂, #Cluster₃, #Cluster₄, and #Cluster₅, we compare our clusters with randomly generated sets. Suppose that $N_i = (a_i, b_i)$ for $i = 1, ..., 10^5$ denotes both the number of Covid-Drug and Clinical-Drug in *i*-th randomly generated sets groups of size n (n=2, 8, 10, 28 and 14). In N_i , a_i and b_i denote the number of Covid-Drug and Clinical-Drug in group of size n, respectively. For nwith sizes of 2, 8, 10, 28 and 14, we define N=(1,1), (1,2) (1,3), (1,1), and (6,8), respectively. Let $X = \{i | N_i > N\}$ for $i = 1, ..., 10^5$, where *X* denotes the number of random sets that performed better than the proposed clusters [31]. The null hypothesis, H_0 , is that our selected drug set of size n is not important. The alternative hypothesis, H_1 , is that our selected drug set of size n is indeed important. We use exceeding value (EV) as $EV = \frac{|X|}{100000}$, where |X|denotes the size of *X*. If $EV < \alpha$ then, we reject H_0 (α is a threshold value that we consider to be 0.05). The values of EV for #Cluster₁, #Cluster₂, #Cluster₃, #Cluster₄,

and $\#Cluster_5$ are reported in Table 5 (These values cause extremely significant results). The EV denotes the percentage of random clusters that perform better than our clusters out of 10^5 random selections. We can conclude that these values are extremely significant and our selected clusters show a better performance than all of these random selections and significantly far from random selection. In Table 6 we also report some details about the proposed clusters. In this table, the number of Covid-Drug, Clinical-Drug, and drugs that are placed in All-Drug groups are reported, respectively.

Evaluation of clusters with respect to proteins as drug targets

For more evaluation, we investigate all of the proteins as drug targets in each cluster. Table 7 shows some details about our selected clusters. In this table, the first row indicates the number of proteins as drug targets in our PPI network. From these proteins, the number of important ones in each cluster that are mentioned as main targets is reported in the second row. These particular targets are the group of proteins from the first row of the table that is placed in set E. The number of human proteins that are targeted with the virus is demonstrated as a set T and reported in the third row. The fourth and fifth rows show the number of these proteins that are targeted through at least one drug in Covid-Drug and Clinical-Drug, respectively. Table 7 reports 10 drugs in #Cluster₃ target 77 human proteins. From these 77 proteins, 10 proteins are reported as particular targets that are identified through our method. This cluster has the highest ratio of the main targets (10/77) in comparison with other clusters. Figure 2 illustrates the number of drug targets for each cluster. Drugs in each cluster can have distinctive and common targets. For example, the union of all targets for two drugs in #Cluster1 contains 28 proteins. From these 28 proteins, 17 of them are common between these two drugs. From these 17 proteins, 1 of them is placed in set *E* that is mentioned as main target.

Table 7 The first row indicates the number of proteins as drug targets in our PPI network

	#Cluster ₁	#Cluster ₂	#Cluster ₃	#Cluster ₄	#Cluster ₅
No. targets	28	37		155	118
No. main targets	2	2	10	10	14
No. targets in T	1	1	1	3	0
No. targets in Covid-Drug	23	18	29	43	47
No. targets in Clinical-Drug	28	32	69	140	107

From these proteins, the number of important ones in each cluster is reported in the second row. The number of human proteins that are targeted with the virus is reported in the third row. The fourth and fifth rows show the number of these proteins that are targeted through at least one drug in Covid-Drug and Clinical-Drug, respectively

Aghdam et al. J Cheminform (2021) 13:70 Page 9 of 14

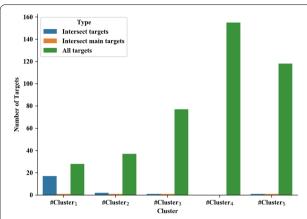


Fig. 2 The blue columns show the common drug targets for each cluster. The orange columns show the number of main targets that are common between drugs in each cluster. The green columns show all of the targets for drugs in each cluster

Figure 2 also shows the number of main targets in each cluster. Each drug in these five clusters has one or multiple main targets. From these main targets three of them are common among all drugs of each clusters. These three main targets are Vascular Endothelial Growth Factor (VEGF)-A, Cytochrome P450 3A4 (CYP3A4), and Prostaglandin-endoperoxide synthase 2 (PTGS2) or Cyclo-oxygenase2 (COX-2), respectively. Despite the lack of evidence for COVID-19, previous research shows that the VEGF family (VEGFs) has a connection with COVID-19. A recent study shows that VEGFs are involved in "cytokine storm" inflammatory response. They claim that these genes may be used as prospective biomarkers for early diagnosis in COVID-19 patients [32]. The VEGFs can also be used for targeted drug delivery in COVID-19 treatment.

The second main target is PTGS2 or COX-2, which has been the subject of many studies on its association with COVID-19 and is a pro-inflammatory enzyme. Some studies showed that the structural proteins of the SARS-CoV family are reported to influence the expression of COX-2 and the increased expression of plasma PGE2 in the blood of SARS-CoV-infected patients. It is also reported that COX-2 plays a crucial role in limiting the anti-viral cytokine response to viral infection. Therefore, the use of an effective COX-2 inhibitor during early viral infection could enhance interferon responses. It might also increase anti-viral immunity [33]. The result of [34] study shows the importance of VEGF-A and COX-2 in relation to COVID-19. In this study, PPI analysis was used to find the hub genes linked to COVID-19 and lung cancer. Among the suggested hub genes, VEGF-A and COX-2 have been confirmed and could be used as biomarkers for COVID-19.

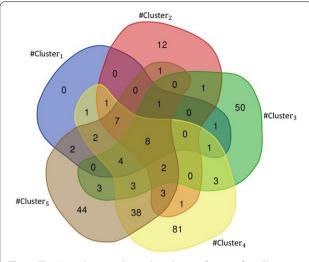


Fig. 3 The Venn diagram shows the relation of targets for #Cluster₁, #Cluster₂, #Cluster₃, #Cluster₄ and #Cluster₅

The next main target is Cytochrome P450 3A4 (CYP3A4). Cytochromes P450 (CYPs) is a superfamily of metabolizing enzymes. The CYP enzymes can be suppressed by an infection-related cytokine increase and inflammation. A recent study demonstrated that, like other viral infections, during the progression of COVID-19 local and systemic inflammation as well as the "cytokine storm" will potentially cause downregulation of the major CYP enzymes including CYP3A4 [35]. A new study proposed that COVID-19 pharmacogenetic studies include CYP3A4 variants [36]. The [33] study shows that CYPs metabolic activity will be surely changed during the SARS-CoV-2 infection in a similar manner, resulting in a pharmacokinetic interaction with the recommended drugs for COVID-19 treatments. In addition, liver involvement in COVID-19 may further complicate this problem, especially for drugs like remdesivir and chloroquine as COVID-19 treatments. Since remdesivir undergoes extensive metabolism by CYPs and chloroquine is also hepatically metabolized, understanding the nature of these drug-disease interactions is highly essential and can affect the therapeutic response of COVID-19 patients.

The Venn diagram in Fig. 3 illustrates the relationship between targets in 5 clusters. Our results show that despite the fact that the drugs in these five clusters are different from each other but they have 8 specific proteins as targets that are jointly targeted by the drugs in all five clusters. In addition, Fig. 3 shows that 64% (50/77) of the proteins as drug targets in #Cluster3 are different from the targets in other clusters, and all proteins as drug targets in #Cluster1 are targeted by at least one drug in the other clusters. In Fig. 4, as an example,

Aghdam et al. J Cheminform (2021) 13:70 Page 10 of 14

we show that the common targets and the total number of targets that are selected through our method with respect to the defined topological features for #Cluster₁.

Evaluation of clusters with respect to related diseases

We have studied the diseases associated with each of the drugs in each cluster based on the information on the Drugbank website. The Venn diagram in Fig. 5 shows the relationship between diseases that are associated with drug targets in each cluster. Figure 5 shows that there is no specific disease that is associated with all clusters. Respiratory Tract Infections and Type 2 Diabetes are two of six diseases that have common targets in#Cluster2 and #Cluster3. Diabetic Macular Edema (DME) is one of two diseases that have common targets in #Cluster3 and #Cluster5. Rheumatoid Arthritis and Ankylosing Spondylitis (AS) are two of twelve diseases that have common targets in #Cluster4 and #Cluster5.

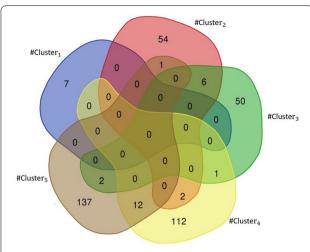
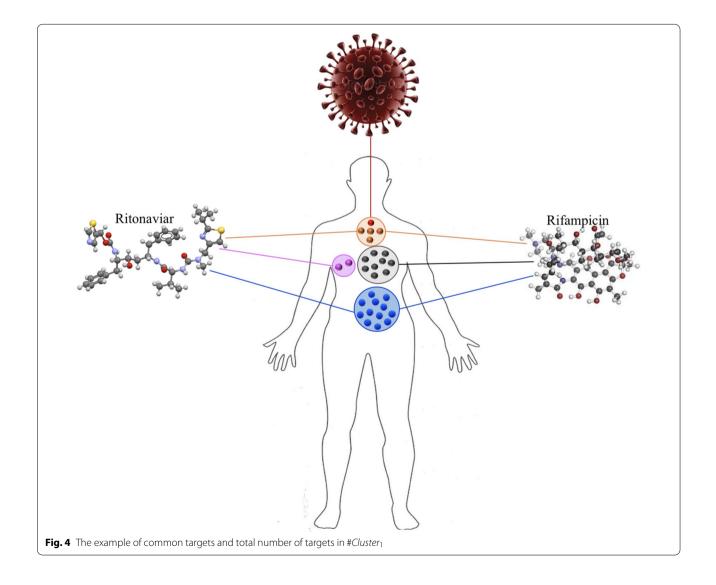


Fig. 5 The relationship between diseases that are associated with drug targets in #Cluster₁, #Cluster₂, #Cluster₃, #Cluster₄ and #Cluster₅



Aghdam et al. J Cheminform (2021) 13:70 Page 11 of 14

Table 8 List of the repositioning candidates, therapeutic category and the supporting published evidence in #Cluster₁

Candidate drug	Therapeutic category	References
Ritonavir	Anti-HIV agents and anti-infective agents	[26]
Rifampicin	Anti-bacterial agents and anti-infective agents	[38]

The drug in Clinical-Drug group is highlighted in italic

Table 9 List of the repositioning candidates, therapeutic category and the supporting published evidence in #Cluster₂

Candidate drug	Therapeutic category	References
Mycophenolate mofetil	Anti-bacterial and anti-infective	[39]
Erythromycin	Anti-bacterial and anti-infective	[40]
Azithromycin	Anti-bacterial and anti-infective	[26]
Cerivastatin	Anticholesteremic agents	[41]
Ezetimibe	Anticholesteremic agents	[42]
Fusidic acid	Anti-bacterial and anti-infective	-
Canagliflozin	Alimentary tract and metabolism	[43]
Letermovir	Antiviral agents	[44]

Drugs in Clinical-Drug group are highlighted in italic

Table 10 List of the repositioning candidates, therapeutic category and the supporting published evidence in #Cluster₃

11 31	9
Therapeutic category	References
Angiogenesis inhibitors and antibodies	[26]
Anti-bacterial and anti-infective agents	[45]
Alimentary tract and metabolism	[46]
Adrenergic agents	[47]
Angiogenesis inhibitors and antibodies	-
Drug delivery systems	[48]
Antineoplastic agents	-
_	-
Heterocyclic compounds	[49]
Allogeneic cultured cell scaffold	[50]
	Angiogenesis inhibitors and antibodies Anti-bacterial and anti-infective agents Alimentary tract and metabolism Adrenergic agents Angiogenesis inhibitors and antibodies Drug delivery systems Antineoplastic agents - Heterocyclic compounds

Drugs in Clinical-Drug group are highlighted in italic

Drug repurposing candidate

In this section, the drug clusters identified with our proposed method have been analyzed to infer some useful drug repurposing candidates. Tables 8, 9, 10, 11 and 12 show repurposing candidates in #Cluster₁, #Cluster₂, #Cluster₃, #Cluster₄, and #Cluster₅, contain 2, 8, 10, 28, and 14 drugs, respectively. Our clustering method enables us to partition the large drug-related network into

Table 11 List of the repositioning candidates, therapeutic category and the supporting published evidence in #Cluster4

Candidate drug	Therapeutic category	References
Aldesleukin	Anti-infective agents	[26]
Dapsone	Anti-infective agents	[51]
Acetaminophen	Central nervous system agents	[52]
Celecoxib	Anti-inflammatory agents	[33]
Rofecoxib	Anti-inflammatory agents	[53]
Valdecoxib	Anti-inflammatory agents	[54]
Diclofenac	Anti-inflammatory agents	[55]
Triamcinolone	Alimentary tract and metabolism	-
Etoposide	Antineoplastic agents	[56]
Phenylbutazone	Anti-inflammatory agents	[57]
Meloxicam	Anti-inflammatory agents	[58]
Chlorphenesin	Central nervous system agents	[59]
Ibuprofen	Anti-inflammatory agents	[26]
Paclitaxel	Antineoplastic agents	-
Drospirenone	Contraceptive agents, female	[60]
Antipyrine	Anti-inflammatory agents	-
Etoricoxib	Anti-inflammatory agents	[53]
Resveratrol	Anti-inflammatory agents	[61]
Nimesulide	Anti-inflammatory agents	[62]
Capsaicin	Sensory system agents	[63]
Parecoxib	Anti-inflammatory agents	[64]
Pomalidomide	Antineoplastic agents	[65]
Cannabidiol	Antidepressive agents	[66]
Loxoprofen	Anti-inflammatory agents	[67]
Dexibuprofen	Anti-inflammatory agents	[68]
Propacetamol	Sensory system agents	[69]
Venetoclax	Antineoplastic agents	-
Nabiximols	Central nervous system agents	_

Drugs in Clinical-Drug group are highlighted in italic

Table 12 List of the repositioning candidates, therapeutic category and the supporting published evidence in #Cluster₅

Candidate drug	Therapeutic category	References
Colchicine	Immunosuppressive agents	[26]
Darunavir	Anti-infective agents	[26]
Dexamethasone	Anti-inflammatory agents	[26]
Methylprednisolone	Anti-inflammatory agents	[26]
Tocilizumab	Antirheumatic agents	[26]
Acetaminophen	Central nervous system agents	-
Chloroquine	Anti-inflammatory agents	[26]
Donepezil	Central nervous system agents	[70]
Clomipramine	Central nervous system agents	[71]
Resveratrol	Anti-inflammatory agents	[72]
Curcumin	Anti-inflammatory agents	[73]
Curcumin sulfate	anti-inflammatory agent	[74]
Baricitinib	Antineoplastic and immunomod- ulating agents	[37]
Lidocaine	Antiarrhythmic agents	[75]

Drugs in Clinical-Drug group are highlighted in italic

Aghdam et al. J Cheminform (2021) 13:70 Page 12 of 14

smaller subgroups and this can simplify the drug repurposing process. Tables 8, 9, 10, 11 and 12 show the mechanism of action, therapeutic category, and supporting published evidence for each drug in #Cluster₁, #Cluster₂, #Cluster₃, #Cluster₄, and #Cluster₅, respectively. From these 62 drugs, 10 of them have previously been proposed as potential drug repurposing candidates for COVID-19 disease.

Discussion and summary

Researchers have been searching for efficient medications to prevent or cure COVID-19 since the first case was discovered in 2019. To advance this goal, we introduced the four steps method. In the first step, the COVID-19 related biological network was constructed and the essential proteins that have a wide range of important functions in the biological network were detected. In the second step, we focused on finding the most effective essential proteins related to COVID-19. To do this, we used two different algorithms to identify the minimum number of proteins that participate in a large number of IBP GO terms and placed them in two distinct sets. Then, we evaluated proteins of these two sets with respect to the number of approved Covid-Drug and Clinical-Drug by them (Table 2). We placed the union of these two sets in the set Cut_{union} and studied set Cut_{union} with respect to the number of IBP GO terms that are disrupted. As a result, the selected proteins can be identified as a suitable candidate set for the COVID-19. It is noticeable that not every essential protein is an appropriate candidate as an essential protein for COVID-19 pathology. Some of these essential proteins are related to the cellular function of the cell and selecting them as drug targets may lead to disruption of cellular function. Considering that, in the third step, we picked candidate proteins directly related to COVID-19 pathology. For the final essential protein selection process in this step, we identified proteins that were associated with underlying diseases such as cardiovascular disease, diabetes, hepatitis, lung, kidney diseases, and various types of cancer. Among 3,002 essential proteins related to COVID-19 in Cutunion, we detected 93 proteins associated with at least four of five underlying mentioned diseases as essential proteins related to COVID-19 pathology (Table 3). We evaluated these proteins with respect to the related pathways with DAVID tools (Table 4). As a result, these selected proteins could be suitable candidates as drug targets for COVID-19 treatment. In the fourth step, multiple informative topological features for drug-target and a PPI network were proposed. Our methods tried to find significant clusters containing appropriate candidate drugs through these features. These features cluster the available experimental unapproved drugs for COVID-19 into five groups (#Cluster₁, #Cluster₂, #Cluster₃, #Cluster₄, and #Cluster₅). These clusters have a significant difference from random clusters (Table 5) and contain a significant number of Covid-Drug and Clinical-Drug (Table 6). We also used three different measures for validating the obtained clusters. The first measure was based on the proteins as drug targets in these clusters, we showed that the proposed clusters have meaningful targets that were known in recent studies as COVID-19 targets (Table 6 and Figs. 2, 3, and 4). The second measure was based on the related diseases that have drugs in our clusters. We found some related diseases like DME and Rheumatoid Arthritis that have drugs in two of our clusters (Fig. 5). The third measure was related to drugs as good candidates for drug repurposing in COVID-19 treatment.

In summary, the main advantage of our method in comparison to other studies was clustering FDAapproved drugs that are related to COVID-19 according to the biological and topological properties of their targets. It can be concluded that partitioning the drugrelated network into smaller networks (clusters) can improve drug repurposing results for clinical trials. In this work, we proposed some good drug candidates as repurposing candidates for COVID-19 treatment. Our results showed that most of our drug candidates were used in clinical trials or suggested in at least one study as suitable drug repurposing candidates (Tables 7-11). Our results also revealed that the proposed informative features recommended some suitable candidate drugs like [37] and Rifampicin [38]. Finally, this study offered powerful network-based informative features for the fast identification of repurposable drugs as a potential treatment for COVID-19. The proposed method can effectively minimize the timing gap between preclinical testing conclusions and clinical results, which is a considerable problem in the fast development of efficient treatment strategies for the emerging COVID-19 outbreak.

Abbreviations

COVID-19: The coronavirus disease 2019; PPI: Protein—protein interaction; IBP: Informative biological processes; DAVID: Database for annotation, visualization, and integrated discovery; BioGRID: Biological general repository for interaction datasets; HIPPIE: Human integrated protein—protein inter-action reference; APID: Agile protein interactomes dataanalyzer; Hint: Homologous interactions; Y2H: yeast-two hybrid; UniProt: Universal protein resource; GO: Gene ontology; EV: Exceeding value; CYP3A4: Cytochrome P450 3A4; VEGF: Vascular endothelial growth factor; CYP3A4: P450 3A4; PTGS2: Prostaglandin-endoperoxide synthase 2; COX- 2: Cyclo-oxygenase2; CYPs: Cytochromes P450.

Acknowledgements

None

Authors' contributions

M. H and G. T developed the methods. M. H, G. T and R. A performed computational and statistical analysis. M. H, G. T and R. A design and wrote the paper. G. T and R. A contributed in writing and editing the manuscript. The authors

Aghdam et al. J Cheminform (2021) 13:70 Page 13 of 14

contributed equally to this work. All authors read and approved the final manuscript.

Funding

No funding to declare.

Availability of data and materials

Datasets and the codes of the algorithms are available in our github repository, [https://github.com/rosaaghdam/Drug-Target.]

Declarations

Ethics approval and consent to participate

None.

Competing interests

The authors declare that they have no competing interests.

Author details

¹School of Biological Sciences, Institute for Research in Fundamental Sciences (IPM), Tehran, Iran. ²Department of Mathematics, Qazvin Branch, Islamic Azad University, Qazvin, Iran. ³Department of Electrical Engineering and Computer Science, KTH Royal Institute of Technology, Stockholm, Sweden. ⁴Science for Life Laboratory, Stockholm, Sweden.

Received: 24 March 2021 Accepted: 6 September 2021 Published online: 20 September 2021

References

- Smith T, Bushek J, LeClaire A, Prosser T (2020) COVID-19 drug therapy. Elsevier 1(1):1–25
- 2. Li J, Zheng S, Chen B, Butte AJ, Swamidass SJ, Lu Z (2016) A survey of current trends in computational drug repositioning. Brief Bioinform
- Habibi M, Taheri G (2021) Topological network based drug repurposing for coronavirus 2019. PLoS One 16(7):0255270
- Bolgár B, Arany A, Temesi G, Balogh B, Antal P, Matyus P (2013) Drug repositioning for treatment of movement disorders: from serendipity to rational discovery strategies. Curr Top Med Chem 13(18):2337–2363
- Forst CV (2010) Host–pathogen systems biology. In: Infectious disease informatics, pp. 123–147
- Kaufmann SH, Dorhoi A, Hotchkiss RS, Bartenschlager R (2018) Hostdirected therapies for bacterial and viral infections. Nat Rev Drug Discov 17(1):35
- Cheng F, Lu W, Liu C, Fang J, Hou Y, Handy DE, Wang R, Zhao Y, Yang Y, Huang J et al (2019) A genome-wide positioning systems network algorithm for in silico drug repurposing. Nat Commun 10(1):1–14
- Alaimo S, Pulvirenti A (2019) Network-based drug repositioning: approaches, resources, and research directions. In: Vanhaelen Q (ed) Computational methods for drug repurposing, vol 1903. Humana Press, New York, pp 97–113
- Bandyopadhyay S, Ray S, Mukhopadhyay A, Maulik U (2015) A review of in silico approaches for analysis and prediction of HIV-1-human protein– protein interactions. Brief Bioinform 16(5):830–851
- Mukhopadhyay A, Maulik U (2014) Network-based study reveals potential infection pathways of hepatitis-c leading to various diseases. PLoS One 9(4):94029
- Cao H, Zhang Y, Zhao J, Zhu L, Wang Y, Li J, Feng Y-M, Zhang N (2017) Prediction of the Ebola virus infection related human genes using protein–protein interaction network. Comb Chem High Throughput Screen 20(7):638–646
- Zhou Y, Hou Y, Shen J, Huang Y, Martin W, Cheng F (2020) Network-based drug repurposing for novel coronavirus 2019-NCOV/SARS-COV-2. Cell Discov 6(1):1–18
- Li X, Yu J, Zhang Z, Ren J, Peluffo AE, Zhang W et al (2021) Network bioinformatics analysis provides insight into drug repurposing for COVID-19. Med Drug Discov 10(1):100090

- Gordon DE, Jang GM, Bouhaddou M, Xu J, Obernier K, White KM, O'Meara MJ, Rezelj VV, Guo JZ, Swaney DL et al (2020) A SARS-COV-2 protein interaction map reveals targets for drug repurposing. Nature 583(1):1–13
- Dick K, Biggar KK, Green JR (2020) Computational prediction of the comprehensive SARS-CoV-2 vs. human Interactome to guide the design of therapeutics. bioRxiv. https://doi.org/10.1101/2020.03.29.014381
- Taheri G, Habibi M, Wong L, Eslahchi C (2013) Disruption of protein complexes. J Bioinform Comput Biol 11(3):1341008
- Habibi M, Khosravi P (2018) Disruption of the protein complexes from weighted complex networks. IEEE/ACM Trans Comput Biol Bioinform 17(1):102–109
- Taheri G, Khonsari A, Entezari-Maleki R, Sousa L (2020) A hybrid algorithm for task scheduling on heterogeneous multiprocessor embedded systems. Appl Soft Comput 91(1):106202
- Luck K, Kim D-K, Lambourne L, Spirohn K, Begg BE, Bian W, Brignall R, Cafarelli T, Campos-Laborie FJ, Charloteaux B et al (2020) A reference map of the human binary protein interactome. Nature 580(7803):402–408
- Chatr-Aryamontri A, Oughtred R, Boucher L, Rust J, Chang C, Kolas NK, O'Donnell L, Oster S, Theesfeld C, Sellam A et al (2017) The biogrid interaction database: 2017 update. Nucleic Acids Res 45(D1):369–379
- 21. Alanis-Lobato G, Andrade-Navarro MA, Schaefer MH (2016) Hippie v2. 0: enhancing meaningfulness and reliability of protein–protein interaction networks. Nucleic Acids Res 45(D1):408–414
- Alonso-López D, Campos-Laborie FJ, Gutiérrez MA, Lambourne L, Calderwood MA, Vidal M, De Las Rivas J (2019) Apid database: redefining protein–protein interaction experimental evidences and binary interactomes. Database 2019(D1):1–10
- 23. Patil A, Nakamura H (2005) Hint: a database of annotated protein–protein interactions and their homologs. Biophysics 1(1):21–24
- Consortium U (2019) Uniprot: a worldwide hub of protein knowledge. Nucleic Acids Res 47(1):506–515
- Ontology G (2019) The gene ontology resource: 20 years and still going strong. Nucleic Acids Res 47(1):330–338
- Wishart DS, Feunang YD, Guo AC, Lo EJ, Marcu A, Grant JR, Sajed T, Johnson D, Li C, Sayeeda Z et al (2018) Drugbank 5.0: a major update to the drugbank database for 2018. Nucleic Acids Res 46(D1):1074–1082
- Kim M, Kim YB (2020) In silico synergistic drug repurposing for combating novel coronavirus (COVID-19) outbreaks. Res Sq. https://doi.org/10. 21203/rs.3.rs-21849/v1
- Oh KK, Adnan M, Cho DH (2020) SARS-COV-2 intervened by nsaids: a network pharmacology approach to decipher signaling pathway and interactive genes. Res Sq. https://doi.org/10.21203/rs.3.rs-111615/v1
- 29. Dai W, Cao D, Zhang W, Wei Y, Ding D, Li B, Gao Y, Zhao L, Jiang Y, Kong X (2020) Integrated bioinformatics analysis reveals key candidate genes and cytokine pathways involved in COVID-19 after rhinovirus infection in asthma patients. Med Sci Monit 26:e928861-1
- Toraih EA, Sedhom JA, Dokunmu TM, Hussein MH, Ruiz EM, Muthusamy K, Zerfaoui M, Kandil E (2021) Hidden in plain sight: the effects of BCG vaccination in the COVID-19 pandemic. J Med Virol 93(4):1950–1966
- Aghdam R, Ganjali M, Zhang X, Eslahchi C (2015) Cn: a consensus algorithm for inferring gene regulatory networks using the sorder algorithm and conditional mutual information test. Mol BioSyst 11(3):942–949
- Veerabathiran R, Ragunath B, Kaviarasan V, Mohammed V, Ahmed SS (2021) Identification of selected genes associated with the SARS-COV-2: a therapeutic approach and disease severity. Bull Natl Res Centre 45(1):1–11
- Baghaki S, Yalcin CE, Baghaki HS, Aydin SY, Daghan B, Yavuz E (2020) Cox2 inhibition in the treatment of COVID-19: Review of literature to propose celecoxib repositioning for randomized controlled studies. Int J Infect Dis 101(1):29–32
- Nana KS, Karuppanan K, Kumar S (2021) Identification of common key genes and pathways between covid-19 and lung cancer by using protein–protein interaction network analysis. bioRxiv. https://doi.org/10. 1101/2021.02.16.431364
- 35. El-Ghiaty MA, Shoieb SM, El-Kadi AO (2020) Cytochrome p450-mediated drug interactions in COVID-19 patients: current findings and possible mechanisms. Med Hypotheses 144:110033
- Fricke-Galindo I, Falfán-Valencia R (2021) Pharmacogenetics approach for the improvement of COVID-19 treatment. Viruses 13(3):413
- 37. Kalil AC, Patterson TF, Mehta AK, Tomashek KM, Wolfe CR, Ghazaryan V, Marconi VC, Ruiz-Palacios GM, Hsieh L, Kline S et al (2020) Baricitinib

- plus remdesivir for hospitalized adults with COVID-19. N Engl J Med 384(9):1–13
- 38. Tripathi V, Pathak Y, Mishra A (2020) Rifampicin may be repurposed for COVID-19 treatment: Insights from an in-silico study. Res Sq. https://doi.org/10.21203/rs.3.rs-22546/v1
- Russell B, Moss C, George G, Santaolalla A, Cope A, Papa S, Van Hemelrijck M (2020) Associations between immune-suppressive and stimulating drugs and novel COVID-19-a systematic review of current evidence. Ecancermedicalscience 14(1022):1–17
- 40. Habibi M, Taheri G, Aghdam R (2021) A SARS-COV-2 (COVID-19) biological network to find targets for drug repurposing. Sci Rep 11(1):1–15
- 41. Rodrigues-Diez RR, Tejera-Muñoz A, Marquez-Exposito L, Rayego-Mateos S, Santos Sanchez L, Marchant V, Tejedor Santamaria L, Ramos AM, Ortiz A, Egido J et al (2020) Statins: could an old friend help in the fight against COVID-19. Br J Pharm 177(21):4873–4886
- 42. Israel A, Schäffer AA, Cicurel A, Feldhamer I, Tal A, Cheng K, Sinha S, Schiff E, Lavie G, Ruppin E (2020) Large population study identifies drugs associated with reduced COVID-19 severity. medRxiv. https://doi.org/10.1101/2020.10.13.20211953
- 43. Kouznetsova VL, Zhang A, Tatineni M, Miller MA, Tsigelny IF (2020)
 Potential COVID-19 papain-like protease PLPRO inhibitors: repurposing
 FDA-approved drugs. PeerJ 8(1):9965
- Cantürk S, Singh A, St-Amant P, Behrmann J (2020) Machine-learning driven drug repurposing for COVID-19. arXiv preprint arXiv:2006.14707
- Singh H, Kakkar AK, Chauhan P (2020) Repurposing minocycline for COVID-19 management: mechanisms, opportunities, and challenges. Expert Rev Anti-infect Therapy 18(10):997–1003
- Tomar PPS, Arkin IT (2020) SARS-COV-2 e protein is a potential ion channel that can be inhibited by gliclazide and memantine. Biochem Biophys Res Commun 530(1):10–14
- 47. Skayem C, Ayoub N (2020) Carvedilol and COVID-19: a potential role in reducing infectivity and infection severity of SARS-COV-2. Am J Med Sci 360(3):300
- Karimi-Zarchi M, Neamatzadeh H, Dastgheib SA, Abbasi H, Mirjalili SR, Behforouz A, Ferdosian F, Bahrami R (2020) Vertical transmission of coronavirus disease 19 (COVID-19) from infected pregnant mothers to neonates: a review. Fetal Pediatr Pathol 39(3):246–250
- Cavasotto CN, Di Filippo JI (2020) In silico drug repurposing for COVID-19: targeting SARS-COV-2 proteins through docking and quantum mechanical scoring. Res Sq. https://doi.org/10.26434/chemrxiv.12110199.v1
- Rivero I, Castresana M, Guala D, Sonnhammer E (2020) Drug repurposing improves disease targeting 11-fold on average. New insights from network module targeting and its application to COVID-19. https://doi. org/10.31219/osfio/4rvaz
- Farouk A, Salman S (2020) Dapsone and doxycycline could be potential treatment modalities for COVID-19. Med Hypotheses 140(1):109768
- Rodríguez-Morales AJ, Cardona-Ospina JA, Murillo-Muñoz MM (2020) Gastroenterologists, hepatologists, COVID-19 and the use of acetaminophen. Clin Gastroenterol Hepatol 18(9):2142–2143
- Organization WH (2020) The use of non-steroidal anti-inflammatory drugs (nsaids) in patients with COVID-19: scientific brief, 19 April 2020. Technical documents
- Dayer MR (2020) Old drugs for jak-stat pathway inhibition in COVID-19. arXiv preprint arXiv:2010.12332
- Kelleni MT (2020) Early use of non-steroidal anti-inflammatory drugs in COVID-19 might reverse pathogenesis, prevent complications and improve clinical outcomes. Biomed Pharmacother 133(1):110982
- Takami A (2020) Possible role of low-dose etoposide therapy for hemophagocytic lymphohistiocytosis by COVID-19. Int J Hematol 112(1):122–124
- Loganathan T, Ramachandran S, Shankaran P, Nagarajan D et al (2020)
 Host transcriptome-guided drug repurposing for COVID-19 treatment: a meta-analysis based approach. PeerJ 8(1):9357

- Farag A, Wang P, Ahmed M, Sadek H (2020) Identification of FDA approved drugs targeting COVID-19 virus by structure-based drug repositioning. ChemRxiv. https://doi.org/10.26434/chemrxiv.12003930.v1
- Mummed Y (2020) Molecular targets for COVID-19 drug development: enlightening nigerians about the pandemic and future treatment. Biosaf Health 2(4)
- Regidor P, Colli E (2021) SARS COVID-19 as an immunothrombotic disease and the potential benefits of a new estrogen-free contraceptive containing drospirenone. Thromb Haemost Res 5(1):1052
- Giordo R, Zinellu A, Eid AH, Pintus G (2021) Therapeutic potential of resveratrol in COVID-19-associated hemostatic disorders. Molecules 26(4):856
- Scalise M, Indiveri C (2020) Repurposing nimesulide, a potent inhibitor of the b0at1 subunit of the SARS-COV-2 receptor, as a therapeutic adjuvant of COVID-19. SLAS DISCOVERY Adv Sci Drug Discov 25(10):1171–1173
- 63. Gonzalez-Paz LA, Lossada CA, Moncayo LS, Romero F, Paz JL, Vera-Villalobos J, Pérez AE, San-Blas E, Alvarado YJ (2020) Theoretical molecular docking study of the structural disruption of the viral 3cl-protease of COVID19 induced by binding of capsaicin, piperine and curcumin part 1: a comparative study with chloroquine and hydrochloroquine two antimalaric drugs. Res Sq. https://doi.org/10.21203/rs.3.rs-21206/v1
- 64. Marak BN, Dowarah J, Khiangte L, Singh VP (2021) Step toward repurposing drug discovery for COVID-19 therapeutics through in silico approach. Drug Dev Res 82(3):374–392
- 65. Sotoudeh E, Sotoudeh H (2020) A hypothesis about the role of fetal hemoglobin in COVID-19. Med Hypotheses 144(1):109994
- Esposito G, Pesce M, Seguella L, Sanseverino W, Lu J, Corpetti C, Sarnelli G (2020) The potential of cannabidiol in the COVID-19 pandemic. Br J Pharmacol 177(21):4967–4970
- 67. Hayakawa J (2021) Favipiravir/loxoprofen/paracetamol: symmetrical drug-related intertriginous and flexural exanthema: case report. React Wkly 1841(1):113
- Siminea N, Popescu V, Martin JAS, Florea D, Gavril G, Gheorghe A-M, Itcus C, Kanhaiya K, Pacioglu O, Popa LI, et al. (2021) Network analytics for drug repurposing in COVID-19. arXiv preprint arXiv:2106.12297
- Li F, Michelson AP, Foraker R, Zhan M, Payne PR (2020) Repurposing drugs for COVID-19 based on transcriptional response of host cells to SARS-COV-2. arXiv e-prints, 2006
- 70. Hashimoto K (2021) Repurposing of CNS drugs to treat COVID-19 infection: targeting the sigma-1 receptor. Eur Arch Psychiatry Clin Neurosci 1(1):1–10
- Nobile B, Durand M, Olié E, Guillaume S, Molès J, Haffen E, Courtet P (2020) Clomipramine could be useful in preventing neurological complications of SARS-COV-2 infection. J Neuroimmune Pharmacol 15(3):347–348
- Marinella MA (2020) Indomethacin and resveratrol as potential treatment adjuncts for SARS-COV-2/COVID-19. Int J Clin Pract 74(9):13535
- Miryan M, Bagherniya M, Sahebkar A, Soleimani D, Rouhani MH, Iraj B, Askari G (2020) Effects of curcumin-piperine co-supplementation on clinical signs, duration, severity, and inflammatory factors in patients with COVID-19: a structured summary of a study protocol for a randomised controlled trial. Trials 21(1):1–2
- Soni VK, Mehta A, Ratre YK, Tiwari AK, Amit A, Singh RP, Sonkar SC, Chaturvedi N, Shukla D, Vishvakarma NK (2020) Curcumin, a traditional spice component, can hold the promise against COVID-19? Eur J Pharmacol 886(1):173551
- Sestili P, Fimognari C (2020) Paracetamol-induced glutathione consumption: is there a link with severe COVID-19 illness? Front Pharmacol 11(1):1597